Genomic DNA Mini Kit (Tissue)

For research use only

Sample: up to 30 mg of tissue (tailsnips, liver, kidney, brain, adipose tissue, earpunches, insects etc.)
Yield: 10-20 µg (0.5 cm of mouse tail, 20 mg of mouse liver), 20-50 µg (20 mg of mouse kidney)
Format: spin column
Time: within 30 minutes
Elution volume: 30-200 µl
Storage: dry at room temperature (15-25ºC)

Geneaid

INTERNATIONAL UKAS QUALITY MANAGEMENT Registration Number 046

CERTIFICATE NO. QAIC/TW/50077

ISO 9001:2008 QMS

Introduction

The Genomic DNA Mini Kit (Tissue) was designed specifically for purifying total DNA (including genomic, mitochondrial and viral DNA) from a variety of tissue and insect samples. The provided micropestle can efficiently homogenize tissue samples to shorten the time in the Lysis Step. Proteinase K and chaotropic salt are used to lyse cells and degrade protein, allowing DNA to be easily bound by the glass fiber matrix of the spin column. Once any contaminants have been removed, using a Wash Buffer (containing ethanol), the purified DNA is eluted by a low salt Elution Buffer, TE or water. The entire procedure can be completed without phenol/chloroform extraction or alcohol precipitation. The purified DNA (approximately 20-30 kb) is suitable for use in PCR or other enzymatic reactions.

Quality Control

The quality of the Genomic DNA Mini Kit (Tissue) is tested on a lot-to-lot basis by isolating genomic DNA from a 20 mg mouse liver sample. The purified DNA (more than 10 µg with an A260/A280 ratio of 1.8-2.0) is quantified with a spectrophotometer and analyzed by electrophoresis.

Kit Contents

Component	GT004	GT050	GT100	GT300
GT Buffer	3 ml	30 ml	30 ml	75 ml
GBT Buffer	4 ml	40 ml	40 ml	75 ml
W1 Buffer	2 ml	45 ml	45 ml	130 ml
Wash Buffer* (Add Ethanol)	1 ml (4 ml)	25 ml (100 ml)	25 ml (100 ml)	50 ml (200 ml)
Proteinase K** (Add ddH_2O)	1 mg (0.1 ml)	11 mg (1.1 ml)	11 mg x 2 (1.1 ml x 2)	65 mg (6.5 ml)
Elution Buffer	1 ml	30 ml	30 ml	75 ml
GS Columns	4	50	100	300
2 ml Collection Tubes	8	100	200	600
Micropestle	4	50	100	300

Order Information

Product	Package Size	Catalogue Number
Genomic DNA Mini Kit (Blood/Cultured Cell)	100/300 preps	GB100/300
Genomic DNA Maxi Kit (Blood/Cultured Cell)	10/25 preps	GDM010/25
Genomic DNA Mini Kit (Tissue)	50/100/300 preps	GT050/100/300
gSYNC™ DNA Extraction Kit	50/100/300 preps	GS050/100/300
Genomic DNA Mini Kit (Plant)	100 preps	GP100
Genomic DNA Maxi Kit (Plant)	10/25 preps	GPM010/25
GENEzol™ DNA Reagent Plant	100/200 rxns	GR100/200
Presto™ Mini gDNA Yeast Kit	100/300 preps	GBY100/300
Presto™ Mini gDNA Bacteria Kit	100/300 preps	GBB100/101/300/301
Geneius™ Micro DNA Extraction Kit	100/300 preps	GMB100/300
Presto™ Buccal Swab gDNA Extraction Kit	100/300 preps	GSK100/300
Presto™ 96 Well Blood gDNA Extraction Kit	4/10 x 96 preps	96GBP04/10
Presto™ 96 Well Plant gDNA Extraction Kit	4/10 x 96 preps	96GPP04/10

*Add absolute ethanol (see the bottle label for volume) to the Wash Buffer prior to initial use
**Add ddH_2O (see the bottle label for volume) to prepare Proteinase K (vortex to dissolve and spin down) and store at 4ºC

Caution

GBT Buffer contains guanidine hydrochloride. During operation, always wear a lab coat, disposable gloves, and protective goggles.

Genomic DNA Mini Kit (Tissue) Functional Test Data

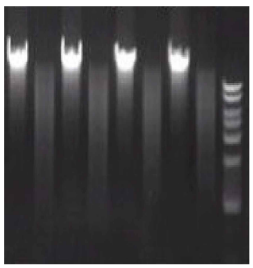

Figure 1. Genomic DNA from a variety of tissue samples was extracted using the Genomic DNA Mini Kit (Tissue). The purified genomic DNA (30-40 kb) was *Eco*RI digested and analyzed by electrophoresis on a 1% agarose gel.
1 = Mouse Liver
2 = Mouse Tail
3 = Fish Muscle
4 = Fruit Fly (Drosophila)
M = Geneaid 1 Kb DNA Ladder

Genomic DNA Mini Kit (Tissue) Protocol

IMPORTANT BEFORE USE

• Add ddH_2O (see the bottle label for volume) to prepare Proteinase K (vortex to dissolve and spin down) and store at 4ºC
• Add absolute ethanol (see the bottle label for volume) to the Wash Buffer prior to initial use
• Additional requirements: microcentrifuge tubes, absolute ethanol, (optional) RNase A (10 mg/ml), ddH_2O

Tissue Dissociation	• Cut up to **30 mg of animal tissue (or 0.5 cm of mouse tail)** then transfer it to a 1.5 ml microcentrifuge tube. NOTE: If tissue has a higher number of cells (e.g. spleen or liver), reduce the starting material to 10 mg. • Use the provided **Micropestle** to grind the tissue to a pulp. • Add **200 µl of GT Buffer** to the tube and homogenize the sample tissue by grinding. • Add **20 µl of Proteinase K** to the sample mixture then shake vigorously and incubate at 60ºC for 30 minutes. NOTE: During incubation, invert the tube every 5 minutes.
Step 1 Lysis	• Add **200 µl of GBT Buffer** then shake vigorously for 5 seconds. • Incubate at 60ºC for at least 20 minutes to ensure the lysate is clear. NOTE: During incubation, invert the tube every 5 minutes. If insoluble material is present following incubation, centrifuge for 2 minutes at 14-16,000 x g then transfer the supernatant to a new 1.5 ml microcentrifuge tube. At this time, preheat the required **Elution Buffer** (200 µl per sample) to 60ºC (for Step 4 DNA Elution). **Optional Step: RNA Degradation** (If RNA free gDNA is required, perform this optional step) • Following 60ºC incubation, add 4 µl of RNase A (10 mg/ml) to the sample lysate then shake vigorously. • Incubate at room temperature for 5 minutes.
Step 2 DNA Binding	• Add **200 µl of absolute ethanol** to the lysate then immediately shake vigorously for 10 seconds. NOTE: If precipitate appears, break it up as much as possible with a pipette. • Place a **GS Column** in a **2 ml Collection Tube**. • Transfer the mixture (including any precipitate) to the **GS Column** then centrifuge at 14-16,000 x g for 2 minutes. • Discard the **2 ml Collection Tube** then transfer the **GS Column** to a new **2 ml Collection Tube**.
Step 3 Wash	• Add **400 µl of W1 Buffer** to the **GS Column** then centrifuge at 14-16,000 x g for 30 seconds. • Discard the flow-through then place the **GS Column** back in the **2 ml Collection Tube**. • Add **600 µl of Wash Buffer** (make sure ethanol was added) to the **GS Column**. • Centrifuge at 14-16,000 x g for 30 seconds. • Discard the flow-through then place the **GS Column** back in the **2 ml Collection Tube**. • Centrifuge for 3 minutes at 14-16,000 x g to dry the column matrix.
Step 4 DNA Elution	Standard elution volume is 100 µl. If less sample is to be used, reduce the elution volume (30-50 µl) to increase DNA concentration. If higher DNA yield is required, repeat the DNA Elution step to increase DNA recovery and the total elution volume to approx. 200 µl. • Transfer the dried **GS Column** to a clean 1.5 ml microcentrifuge tube. • Add **100 µl of pre-heated Elution Buffer** or TE to the CENTER of the column matrix. • Let stand for at least 5 minutes to ensure the **Elution Buffer** or TE is completely absorbed. • Centrifuge at 14-16,000 x g for 30 seconds to elute the purified DNA.

Troubleshooting

Problem	Possible Reasons/Solution
Clogged Column	**Too much tissue was used** • If using more than 30 mg of tissue, separate into multiple tubes. **Sample tissue was not lysed completely** • Add additional Proteinase K and extend the incubation time in the Lysis Step. • Following the Lysis Step, centrifuge for 2 minutes at 14-16,000 x g to remove sample debris. Transfer the supernatant to a new microcentrifuge tube and proceed with the DNA Binding Step. **Precipitate was formed at DNA Binding step** • Reduce the sample material. • Following ethanol addition, break up any precipitate as much as possible prior to loading GS Column.
Low Yield	**Sample tissue was not lysed completely** • Add additional Proteinase K and extend the incubation time in the Lysis Step. **Column was clogged at DNA Binding step** • Following the Lysis Step, remove the insoluble debris by centrifugation. • Prior to loading the column, break up the precipitate in the ethanol-added lysate. **Incorrect DNA Elution Step** • Ensure that the Elution Buffer or TE is added to the center of the GS Column matrix and is absorbed completely. **Incomplete DNA elution** • Elute twice to increase the DNA recovery.
Eluted DNA does not perform well in downstream applications	**Residual ethanol contamination** • Following the Wash Step, dry the GS Column by centrifuge at 14-16,000 x g or incubate at 60ºC for 5 minutes. **RNA/Protein contamination** • Perform optional RNA Degradation step/reduce the sample amount. **Genomic DNA was degraded** • Use fresh samples or freeze fresh samples in liquid nitrogen immediately and store at -80ºC.