University of South Bohemia in České Budějovice Faculty of Science

Causes of endemic radiation in the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini)

RNDr. Thesis

Pável Fortunato Matos Maraví, MSc

České Budějovice 2015 MATOS-MARAVÍ P.F. 2015. Causes of endemic radiation in the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini) [RNDr. Thesis] – 18 p., Faculty of Science, University of South Bohemia, České Budějovice, Czech Republic.

Annotation

In this study, the systematics, historical biogeography, and phylogenetic diversification dynamics of the only satyrine butterfly group occurring in the West Indies were investigated. Inferences on the evolutionary history of the butterfly genus *Calisto* (Lepidoptera: Nymphalidae: Satyrinae) were examined using molecular phylogenetics and molecular clock methods, on a dataset consisting of about 82% of its total extant species diversity and the DNA sequences of six gene markers.

Declaration [in Czech]

Prohlašuji, že svoji rigorózní práci jsem vypracoval samostatně pouze s použitím pramenů a literatury uvedených v seznamu citované literatury.

Prohlašuji, že v souladu s § 47b zákona č. 111/1998 Sb. v platném znění souhlasím se zveřejněním své rigorózní práce, a to [v nezkrácené podobě – v úpravě vzniklé vypuštěním vyznačených částí archivovaných Přírodovědeckou fakultou] elektronickou cestou ve veřejně přistupné částí databáze STAG provozované Jihočeskou univerzitou v Českých Budějovicích na jejích internetových stránkách, a to se zachováním mého autorského práva k odevzdanému textu této kvalifikační práce. Souhlasím dále s tím, aby toutéž elektronickou cestou byly v souladu s uvedeným ustanovením zákona č. 111/1998 Sb. zveřejněny posudky školitele a oponentů práce i záznam o průběhu a výsledku obhajoby kvalifikační prací Theses.cz provozovanou Národním registrem vysokoškolských kvalifikačních prací a systémem na odhalování plagiátů.

V Českých Budějovicích, 19.10.2015

Podpis studenta

Author's contribution

As the first author, I lead most of data generation, research and writing. Following is the detailed list of my roles in this study:

- The initial research plan was conceived along with Dr. Niklas Wahlberg (last author in the study).

- I conducted the laboratory work by isolating DNA from butterflies (along with Carlos Peña and Rayner Núñez) and by sequencing DNA gene fragments.

- I lead the entire data analyses and research discussion. I analyzed the data using phylogenetic and molecular clock methods, likelihood models to reconstruct ancestral geographical ranges and to estimate diversification rates.

– I wrote the first draft of the manuscript and co-authors actively contributed to the final published version of the article.

Financial support and acknowledgements

Field work was covered by each author's institution as well as other grants such as the National Geographic Society (grant #5717-96 to Dr. Andrei Sourakov). Laboratory work expenses were covered by the Finnish Kone Foundation's grant awarded to Dr. Niklas Wahlberg. Personal grants were awarded to me during the development of this study by the Finnish Oskar Öflunds and the University of Turku Foundations, as well as by the University of South Bohemia Grant Agency (GAJU grant 156/2013/P).

I would like to acknowledge my Master thesis supervisor Dr. Niklas Wahlberg for his constant support throughout the length of my stay in Finland, as well as other co-authors of this study for their significant contribution to the quality of the published article. Peer-review referees, including Andrew V.Z. Brower, Fabien L. Condamine and anonymous reviewers, are acknowledged for their constructive comments. I thank a number of researchers that helped me with the analyses and provided further scientific discussion, among them Nick J. Matzke, Rampal S. Etienne, and Jana Smrčková. Finally, I cannot thanks enough to all the support and trust deposited in my person by my parents, my brother, family in Peru, and my new family in Czech Republic, especially to Markéta Aubrechtová.

Article published in BMC Evolutionary Biology. Impact Factor of 3.37

Matos-Maraví P., Núñez Aguila R., Peña C., Miller J.Y., Sourakov A., Wahlberg N. 2014. Causes of Endemic radiation in the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini). *BMC Evol Biol* **14**: 199.

© BioMed Central, BMC Series.

RESEARCH ARTICLE



Open Access

Causes of endemic radiation in the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini)

Pável Matos-Maraví^{1,2*}, Rayner Núñez Águila³, Carlos Peña¹, Jacqueline Y Miller⁴, Andrei Sourakov⁴ and Niklas Wahlberg¹

Abstract

Background: *Calisto* is the largest butterfly genus in the West Indies but its systematics, historical biogeography and the causes of its diversification have not been previously rigorously evaluated. Several studies attempting to explain the wide-ranging diversity of *Calisto* gave different weights to vicariance, dispersal and adaptive radiation. We utilized molecular phylogenetic approaches and secondary calibrations points to estimate lineage ages. In addition, we used the dispersal-extinction-cladogenesis model and Caribbean paleogeographical information to reconstruct ancestral geographical distributions. We also evaluated different models of diversification to estimate the dynamics of lineage radiation within *Calisto*. By understanding the evolution of *Calisto* butterflies, we attempt to identify the main processes acting on insular insect diversity and the causes of its origin and its maintenance.

Results: The crown age of *Calisto* was estimated to the early Oligocene (31 ± 5 Ma), and a single shift in diversification rate following a diversity-dependent speciation process was the best explanation for the present-day diversity found within the genus. A major increase in diversification rate was recovered at 14 Ma, following geological arrangements that favoured the availability of empty niches. Inferred ancestral distributional ranges suggested that the origin of extant *Calisto* is in agreement with a vicariant model and the origin of the Cuban lineage was likely the result of vicariance caused by the Cuba-Hispaniola split. A long-distance dispersal was the best explanation for the colonization of Jamaica and the Bahamas.

Conclusions: The ancestral geographical distribution of *Calisto* is in line with the paleogeographical model of Caribbean colonization, which favours island-to-island vicariance. Because the sister lineage of *Calisto* remains ambiguous, its arrival to the West Indies remains to be explained, although, given its age and historical biogeography, the hypothesized GAARlandia land bridge might have been a plausible introduction route from continental America. Intra-island radiation caused by ecological innovation and the abiotic creation of niche spaces was found to be the main force shaping *Calisto* diversity and island endemism in Hispaniola and Cuba.

Keywords: Caribbean, Ecological limits, Historical biogeography, Intra-island diversification, Island-island vicariance, Lepidoptera, Molecular phylogeny

* Correspondence: pavelm14@gmail.com

²School of Biological Sciences, University of South Bohemia and Institute of Entomology, Biology Centre AS CR, CZ-37005 Ceske Budejovice, Czech Republic

Full list of author information is available at the end of the article



© 2014 Matos-Maraví et al.; licensee BioMed Central Ltd. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly credited. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.

¹Laboratory of Genetics, Department of Biology, University of Turku, FI-20014 Turku, Finland

Background

The Caribbean has been an important model system for studying biotic over-water dispersal from continents and island colonization [1-4], as well as vicariance [5,6] as mechanisms for the origin of diversity, and within-island diversification as mediators of species richness and endemism [7,8]. The geological evolution of the region has certainly had a strong influence on the diversification of species there, and a general understanding of the former is crucial to an understanding of the latter.

The larger islands of the Greater Antilles (i.e. Cuba, Hispaniola, Jamaica and Puerto Rico) were repeatedly submerged until the mid/late Eocene (~40 Ma) [3,6]. A general terrene uplift is likely to have occurred during the mid-Eocene and the early Oligocene (~45-30 Ma), and some authors hypothesized the existence of a land corridor connecting northern South America to the Greater Antilles and subaerial Aves Ridge (GAARlandia, ~35-33 Ma) [6,9], although this is still under debate [10]. Hispaniola and Puerto Rico were physically connected until the formation of the Mona Passage, becoming fully separated during the late Oligocene to early Miocene (~30-20 Ma) [11,12]. Later, during the early to mid-Miocene, the aerial connection between eastern Cuba and northern Hispaniola was interrupted by the expansion of the Windward Passage (~17-14 Ma) [6,13].

Northern and southern Hispaniola paleoislands collided in the mid Miocene (ca. 15-10 Ma) [6,14,15], triggering the initial uplift of south-western Hispaniolan mountains as well as the significant elevation of the Cordillera Central [16,17]. Multiple marine incursions in the Cul-de-Sac/ Enriquillo depression repeatedly separated northern and southern paleoislands until the Plio-Pleistocene (~2.5 Ma) [15,18]. Cuba was fragmented into distinct land blocks comprising the current western, central and eastern parts of the island until the late Miocene, when the closure of the Havana-Matanzas Channel began some 8–6 Ma [6]. On the other side, Jamaica was continuously submerged until ca. 12 Ma [19]. The western Jamaica land block was temporally aerial and connected to Central America during the early to mid-Eocene [6,20], whereas eastern Jamaica (Blue Mountains Block) was apparently connected to GAARlandia through the southern peninsula of Hispaniola during ~35-33 Ma [4,6]. Most Bahamian shallows and keys were repeatedly submerged during the Pliocene and Pleistocene (~4-0.5 Ma) [21].

The butterfly genus *Calisto* (Nymphalidae, Satyrinae, Satyrini) is the only satyrine group occurring in the Caribbean region [22,23]. This genus exhibits remarkable radiation and significantly contributes to the high butterfly endemism seen in the region [24,25]. The genus *Calisto* comprises 44 described species, all geographically restricted to single islands [23,26-29]; 11 distributed in Cuba, 1 in Puerto Rico, 1 in Anegada Island, 1 in Jamaica, 2 in the

Bahamas and the remaining 28 species occurring in Hispaniola. Molecular data has given insight into the cryptic condition of several taxa in Hispaniola [27], as well as assisted in determining the phylogenetic relationships of Cuban taxa [28].

Even though the monophyly of the genus appears to be clear [27], its position within the taxonomic tribe Satyrini has not been resolved. Morphological studies classify *Calisto* within the subtribe Pronophilina [23], closely related to the Neotropical genus *Eretris* [30,31]. However, this has not been corroborated at the molecular level [32]. Certain morphological similarities have even led some authors to propose African affinities with, for instance, the subtribe Ypthimina (Satyrini) [33] and the satyrine tribe Dirini [34,35].

Regardless of the phylogenetic position of *Calisto*, a continental origin of the genus is the most plausible explanation, as no other extant satyrine butterflies with the potential of being a closely related group are found in the Greater Antilles; thus, its ancestors would have necessarily arrived to the Caribbean from the nearby American continent [31,33,36]. Once *Calisto* colonized the Greater Antilles, further differentiation by vicariance [31,37], within-island diversification [28,36] or adaptive radiation [27] might have shaped the evolution of these butterflies.

In this study, we aim to elucidate the phylogenetic affinities and to identify the main drivers of the diversification and distribution of *Calisto* by using a secondarily calibrated molecular phylogeny. We also aim to reconstruct the historical biogeography of *Calisto* and to evaluate possible changes in diversification rates throughout the evolution of the genus. Intra-island differentiation appears to be an important factor for the radiation of these butterflies, a phenomenon observed in other Caribbean animal lineages [2,4,9,38-40]. However, even though rapid diversification driven by ecological evolution is plausible explanation considering the diversity of Caribbean habitats, niche saturation and island size may have imposed diversification limits [38] which could have restricted the diversity and geographical distribution of *Calisto*.

Results

Systematics and divergence dates of Calisto

Our phylogenetic inferences using single gene datasets are congruent with the combined analyses, recovering the main clades within *Calisto* (Additional file 1). Moreover, the combined analyses were consistent regardless of the method used and the partitioning strategy (Figure 1). A summary of the dataset properties is presented in Table 1.

Calisto nubila split early in the evolution of the genus, becoming an old and separate entity. The lineage did not apparently diversify further within Puerto Rico, al-though *C. anegadensis* on Anegada Island might have been derived from it based on morphological similarities

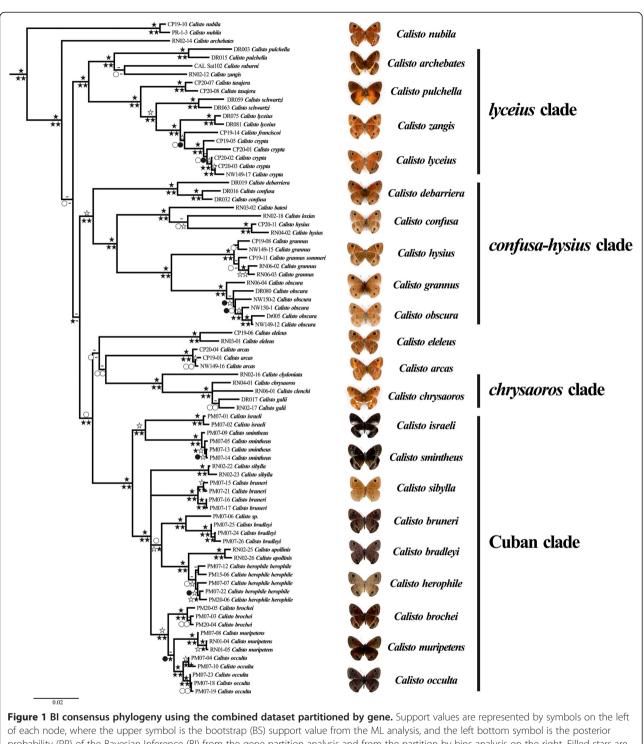


Figure 1 BI consensus phylogeny using the combined dataset partitioned by gene. Support values are represented by symbols on the left of each node, where the upper symbol is the bootstrap (BS) support value from the ML analysis, and the left bottom symbol is the posterior probability (PP) of the Bayesian Inference (BI) from the gene partition analysis and from the partition-by-bins analysis on the right. Filled stars are strong support values of 0.95-1.00 and 90–100 for PP and BS respectively, stars are 0.85-0.94 and 75–89, filled circles are 0.75-0.84 and 65–74 whereas circles are 0.50-0.74 and 50–64. Dashes (–) are unresolved nodes on each analysis. Branch lengths represent expected substitutions/site estimated in the BI analysis.

[26]. Three main monophyletic groups from Hispaniola are identified: the *lyceius-*, the *confusa-hysius* and the *chrysaoros* clades. *Calisto zangis* from Jamaica is likely to have had Hispaniolan ancestors as it belongs to the

"lyceius clade". The monophyletic group consisting of Cuban and Bahamian *Calisto* is closely related to Hispaniolan lineages such as *C. arcas* and the *"chrysaoros* clade", although the relationship among them was not

Partitions	Base pairs	Variable	Informative	Subs. model	<i>m</i> (ratemult)	<i>Alpha</i> (Г shape)	Tree likelihood
Gene strategy							
COI	1487	651	486	GTR + G	1.569	0.245	-18584.5
CAD	850	319	218	HKY + G	0.749	0.282	-4766.8
EF1a	1240	432	290	GTR + G	0.768	0.222	-21045.8
GAPDH	691	264	196	GTR + G	0.864	0.232	-13990.5
RPS5	617	228	183	GTR + G	0.694	0.198	-11920.2
WINGLESS	400	191	130	K80 + G	0.848	0.34	-8617.8
"Bin" strategy							
BIN1	2727	-	-	F81	0.0002	-	-
BIN2-BIN10	652	-	-	GTR	0.57	-	-
BIN11	1269	-	-	GTR + G	1.457	1.601	-
BIN12	637	-	-	GTR + G	4.828	4.389	-

 Table 1 Partition strategies for phylogenetic analyses of the combined dataset

Number of variable and phylogenetically informative sites in our *Calisto* data are shown by gene partition. Substitution model was selected based on BIC calculations in jModelTest [41]. Rate multiplier (*m*) and Gamma-shape (*alpha*) parameters are from BI whereas the *tree likelihood* for each gene partition are from the dating analysis using normal distribution for the calibration points and the birth-death process. Other dating analyses have similar values as shown in *tree likelihood*.

resolved with strong support (Figures 1 and 2). A revised checklist of the genus *Calisto* is presented in Table 2.

The genus Calisto was not recovered within any valid Satyrini subtribes. Instead, our BEAST reconstructions place it sister to all sampled subtribes except Euptychiina with low support values (posterior probability around 0.60-0.65) (Additional file 1). The exclusion of the genus Euptychia (which apparently caused long branch attraction in a different dataset [32]) only increases the support for such a placement to moderate values (around 0.80-0.85). Using birth-death/Yule and normal/uniform as tree processes and calibration distributions respectively does not result in any significant difference in both tree topology and estimated ages (Additional file 1, Figure 2). Height posterior distributions displayed normally whereas summarizing the trees as means or medians height showed no significant difference. The crown age of *Calisto* is inferred at 31 Ma (±5 Ma) in all cases except when Yule process and the calibration normal distribution are used together, in which case the estimate is at 33 Ma (\pm 7 Ma).

Historical biogeography reconstruction

There was no statistical difference in the global likelihood between the non-time-stratified analyses NS0 and NS1 (Table 3). Excluding unlikely area connections (NS1), resulted in a Puerto Rico-northern Hispaniola (PR-nH) distribution on the crown node of *Calisto*, whereas NS0 equally preferred PR along with both nH and sH (southern Hispaniola) (Table 4, Figure 3). Similarly, NS0 and NS1 were unable to discern between dispersal and vicariance for the origin of Cuban *Calisto*. The time-stratified analysis TS1 favoured vicariance over dispersal in all cases and TS2 inferred a PR-sH origin of *Calisto* and vicariance for the origin of Cuban diversity. However, TS2 analysis did not improve the global likelihood of the inference over TS1. Root optimizations significantly favoured a PR-sH distribution and vicariance as the cause of the Cuban clade split from its sister Hispaniolan lineages.

The estimation of the parameter *j* (founder-event speciation) significantly improved the DEC models. The global likelihood of TS1 was improved using BioGeo-BEARS because, in contrast to Lagrange C++, we were able to constrain the area-connectivity through time slices. NS1-*j* preferred dispersal in critical nodes, i.e. the colonization of Jamaica, Cuba and the Bahamas, as well as a widespread origin of Calisto (PR-nH-sH) followed by vicariance. However, from all four models used in BioGeoBEARS, Akaike weights and likelihood-ratio test (LRT) suggested that TS1-*j* had a higher probability of being the best model, followed by TS1. Dispersal to Jamaica and the Bahamas are fully recovered in both TS1 and TS1-*j* from BioGeoBEARS, whereas vicariance is favoured as an explanation of the origin of Cuban Calisto only in TS1 analysis.

Diversification processes within Calisto

The ΔAIC_{RC} critical value for small phylogenies, as estimated in *laser*, is 4 [47]. The observed value for *Calisto* is significantly higher than this threshold (ΔAIC_{RC} = 13), favouring a rate-variable diversification model. However, there was no statistical difference between the rate-variable models Yule-3-rates (Y3r) and the logistic density-dependent (DDL) ($\Delta AIC_{Y3r} - DDL = 3.8$). The diversification of the main *Calisto* tree excluding the Cuban lineage also fits the rate-variable process ($\Delta AIC_{RC} = 11$) better, but there was not strong preference among DDL, Yule-2, and -3-rates ($\Delta AIC_{DDL} - _{Y3r} = 2.0$; $\Delta AIC_{DDL} - _{Y2r} = 3.8$) (Table 5).

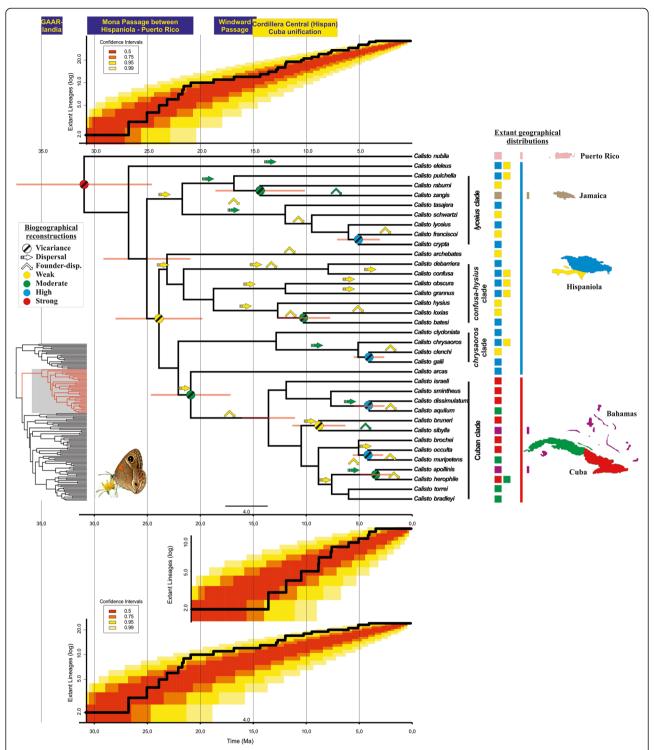


Figure 2 Dated phylogram and a consensus biogeographical history. The ultrametric tree is scaled in Ma. Symbols on each critical node/ branch are depicted as the most likely scenarios: vicariance, dispersal or founder-event. Colours on each symbol represent the level of support. Horizontal bars on nodes represent 95% credibility intervals. The phylogeny in the bottom left is the Satyrini tree, with the *Calisto* clade showing in red. Extant distributions of *Calisto*, following the subdivision of the Greater Antilles, are represented by coloured squares. The main geological events through time are depicted on top of the figure following the time scale in Ma. Lineage Through Time (LTT) plot of extant *Calisto* diversity (log scale) vs. time (Ma) is shown above the phylogeny, whereas the LTT of the Cuban clade is below the tree and the LTT of the Hispaniolan lineages is in the bottom of the figure. LTT plots follow the time scale of the phylogeny in Ma. Confidence intervals for LTT are displayed as coloured ranges.

Table 2 Revised checklist of the genus Calisto
(Lepidoptera: Nymphalidae: Satyrinae: Satyrini)

Island	Proposed species for Calisto	Classification within Calisto
Anegada	Calisto anegadensis	Related to <i>Calisto nubila.</i> Reference [26]
Bahamas	Calisto apollinis	Cuban clade 2
Bahamas	Calisto sibylla	Cuban clade 2
Cuba	Calisto aquilum	Cuban clade 1
Cuba	Calisto bradleyi	Cuban clade 2
Cuba	Calisto brochei	Cuban clade 2
Cuba	Calisto bruneri	Cuban clade 2
Cuba	Calisto dissimulatum	Cuban clade 1
Cuba	Calisto herophile	Cuban clade 2
Cuba	Calisto israeli	Cuban clade 1
Cuba	Calisto muripetens	Cuban clade 2
Cuba	Calisto occulta	Cuban clade 2
Cuba	Calisto smintheus	Cuban clade 1
Cuba	Calisto torrei	Cuban clade 2
Hispaniola (North and South)	Calisto chrysaoros	chrysaoros clade
Hispaniola (North and South)	Calisto confusa	confusa-hysius clade
Hispaniola (North and South)	Calisto eleleus	Incertae sedis
Hispaniola (North and South)	Calisto grannus	confusa-hysius clade
Hispaniola (North and South)	Calisto obscura	confusa-hysius clade
Hispaniola (North and South)	Calisto pulchella	<i>lyceius</i> clade
Hispaniola (North)	Calisto ainigma	Related to <i>Calisto eleleus.</i> Reference [42]
Hispaniola (North)	Calisto arcas	Incertae sedis
Hispaniola (North)	Calisto batesi	confusa-hysius clade
Hispaniola (North)	Calisto clydoniata	chrysaoros clade
Hispaniola (North)	Calisto crypta	lyceius clade
Hispaniola (North)	Calisto debarriera	confusa-hysius clade
Hispaniola (North)	Calisto galii	chrysaoros clade
Hispaniola (North)	Calisto lyceius	lyceius clade
Hispaniola (North)	Calisto neochma	Related to <i>Calisto</i> <i>clydoniata</i> . Reference [43]
Hispaniola (North)	Calisto tasajera	lyceius clade
Hispaniola (North)	Calisto wetherbeei	Related to <i>Calisto</i> <i>archebates</i> . Reference [44]
Hispaniola (South)	Calisto archebates	Incertae sedis
Hispaniola (South)	Calisto clenchi	chrysaoros clade
Hispaniola (South)	Calisto franciscoi	lyceius clade
Hispaniola (South)	Calisto hysius	confusa-hysius clade
Hispaniola (South)	Calisto loxias	confusa-hysius clade

Table 2 Revised checklist of the genus *Calisto* (Lepidoptera: Nymphalidae: Satyrinae: Satyrini) (*Continued*)

Hispaniola (South)	Calisto pauli	Related to <i>Calisto hysius</i> (C. <i>herophile & C. sibylla?</i>). Reference [45]
Hispaniola (South)	Calisto raburni	lyceius clade
Hispaniola (South)	Calisto schwartzi	lyceius clade
Hispaniola (South)	Calisto thomasi	Related to <i>Calisto confusa.</i> Reference [45]
Hispaniola (South)	Calisto tragius	Related to <i>Calisto eleleus.</i> Reference [46]
Hispaniola (South)	Calisto woodsi	Related to <i>Calisto pauli.</i> Reference [45]
Jamaica	Calisto zangis	<i>lyceius</i> clade
Puerto Rico	Calisto nubila	Puerto Rican lineage

Each island and its fauna is shown according to the phylogenetic relationships presented in this study. Hispaniola is subdivided in northern and southern paleoislands. The eight species that were not included in this work are listed with their putative sister taxa.

From all rate-variable models in *DDD*, only those with a whole *Calisto* shift under diversity-dependent process are preferred with Akaike weights higher than 0.1. One single shift in the *K* parameter ("clade-level carrying capacity") at 14 Ma fits 2–3 times better than shifts in *K* along with speciation or extinction rates. The decoupling of parameters for the Cuban taxa alone from the main *Calisto* tree was not enough to explain the radiation of the genus. Cuban and Hispaniolan taxa analyzed separately did not have constant diversification rates; rates changed possibly due to increased speciation, diversitydependence processes, or a combination of both (Akaike Weights were unable to discern among models). Including the number of missing taxa into the models when possible did not affect the recovered estimations (Table 6).

Discussion

Colonization of the Greater Antilles by Calisto

The variability in our dataset (39% and 28% of all characters were variable and phylogenetically informative respectively; Table 1) is similar to previous inter-generic studies in Nymphalidae [32,48,49], but relatively higher than intra-generic studies [50,51]. The genus Calisto is most likely a "relict" satyrine group that might have colonized the Greater Antilles during the uplift of GAARlandia (~35-33 Ma) [32]. Our dating estimates, indeed, confirm that it is an old and independent lineage, and its crown age $(31 \pm 5 \text{ Ma})$ provides evidence in support of the GAARlandia origin. Previous attempts to date the diversification of Calisto were done based only on a pairwise substitution rate for mitochondrial evolution [27]. This latter study deduced younger ages (4–8 Ma) but did not actually carry out a timing of the divergence analysis, rather they only calculated pairwise genetic distances

Non-stratified	Global in-likelihood	d	е	j	Stratified	Global in-likelihood	D	е	j
Lagrange C++					Lagrange C++				
*NS0	-73.5273	0.2700	0.0102	-	*TS1	-88.2411	0.6363	0.0003	-
*NS1	-73.8920	0.3872	0.0146	-	TS2	-93.1298	0.7224	0.0005	-
Root optimization					Root optimization				
*NS1_PR-sH	-72.1124	0.4854	0.0053	-	*TS2_PR-sH	-94.3276	0.6718	0.0005	-
NS1_PR-nH	-74.5323	0.3517	0.0114	-	TS2_PR-nH	-97.4394	0.6162	0.0025	-
NS1_PR-nH-eC	-76.5541	0.3825	0.0118	-	TS2_PR-nH-eC	-97.6103	0.6174	0.0017	-
NS1_PR-sH-eC	-76.6322	0.3313	0.0086	-	TS2_PR-nH-sH	-97.8598	0.5802	0.0019	-
NS1_PR-nH-sH	-77.1248	0.3048	0.0083	-	TS2_PR-sH-eC	-98.4153	0.5613	0.0001	-
NS1_nH	-78.4231	0.3522	0.0141	-	TS2_sH	-99.4647	0.7906	0.0050	-
NS1_PR	-80.0185	0.3219	0.0059	-	TS2_nH	-101.7850	0.6684	0.0089	-
NS1_sH	-80.2485	0.2874	0.0082	-	TpS2_PR	-103.8450	0.6690	0.0063	-
BioGeoBEARS DEC mod	lel				BioGeoBEARS DEC mode	1			
*NS1 <i>-j</i>	-78.7492	0.0020	0.0000	0.0709	*TS1 <i>-j</i>	-63.8944	0.0581	0.0042	0.582
NS1	-97.9848	0.0054	0.0069	-	TS1	-76.7373	0.1211	0.0098	-

ad alabal likalihaada ay aada

The best models from each type of analyses are highlighted in bold text and marked with an asterisk (*). Parameter d is the rate of "dispersal" or range expansion, e is the rate of "extinction" or range contraction, and j is the relative weight of jump dispersal. j is cladogenetic, and d and e are anagenetic processes. Modelcomparison between the BioGeoBEARS models resulted in Akaike weights favouring TS1-j with a relative probability of 0.999 of it being the best model. Similarly, LRT between TS1 and TS1-j, the two best models, rejected TS1 as the null model with fewer parameters with p-value of 4.02e⁻⁰

with Kimura 2-parameter without an adequate model testing.

It is not the first time that GAARlandia is invoked to explain butterfly geographic range expansion. It is the case for the nymphaline subtribe Phyciodina [52], the satyrine subtribe Pronophilina [32], and certain lineages within the papilionid tribe Troidini [53]. The idea of indirect overwater dispersal by "hitch hiking" on hurricanes or flotsams rafts seems unlikely. Adult butterflies respond to incoming bad weather by taking refuge [36] whereas a high mortality of eggs, larvae and pupae is observed when they are exposed to marine water [54]. Calisto, when compared to most other butterflies, are rather sedentary, and hence the direct and indirect dispersal capabilities of Calisto make a dispersalist model less likely.

According to Iturralde-Vinent's vicariance model [6], after GAARlandia, Hispaniola and Puerto Rico split around 20-30 Ma, whereas in our study, extant Calisto species in both islands have their most recent common ancestor at 27 ± 5 Ma. Furthermore, the Cuban clade branched off from a Hispaniola lineage at 21 ± 4 Ma, but did not apparently diversify into any extant Calisto until 14 ± 3 Ma, while the last aerial connection between blocks of Hispaniola and Cuba existed until 14-17 Ma. Therefore, the evolution of *Calisto* is better explained by the main predictions of the Caribbean paleogeographical model of colonization rather than the stochastic dispersalist scenario.

The inclusion of Jamaica into the vicariance model is less supported by the paleogeographical reconstructions,

although a remote connection between the Blue Mountains block with GAARlandia has not been discarded [4,6]. The sole extant Jamaican Calisto split from its Hispaniolan sister taxa at 14 ± 4 Ma. At that time, large portions of Jamaica began to uplift and the entire island remained above water afterwards, and hence the colonization of Jamaica by rare long-distance dispersal events is the most likely explanation for the origin of the endemic sole species found there, Calisto zangis.

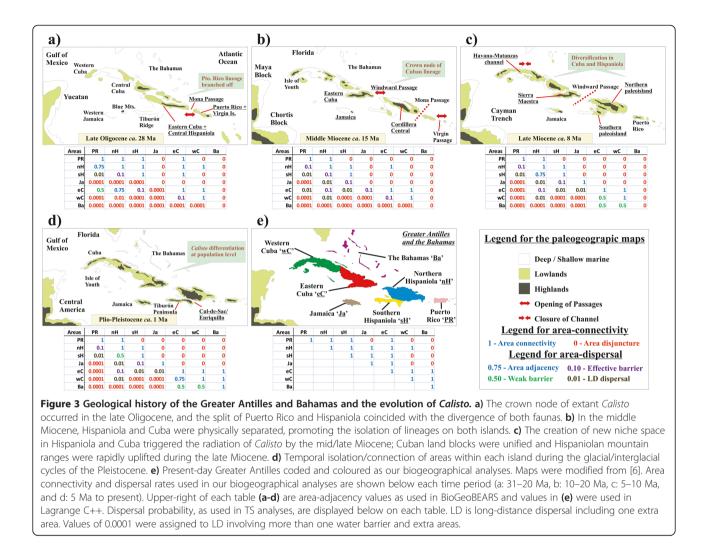
Historical biogeography of Calisto

Biogeographical reconstructions were significantly improved when we constrained dispersal probability and area-connectivity following the paleogeographic history of the Caribbean. Moreover, we found for the first time, statistical support for long-distance dispersal in the colonization of the Bahamas and Jamaica by estimating a founder-event parameter using a more general DEC model. Vicariance was recovered as the main explanation for the first diversification event of Calisto, although we did not find a fully supported dispersal/vicariance origin for the Cuban clade. Whereas NS1 and TS1 in BioGeoBEARS and TS2 did significantly recover vicariance, other analyses did not favour either dispersal nor vicariance. This could be due primarily to, first, the assumptions made by the models and, second, the different approaches to node reconstruction. In the first case, vicariance is favoured when incorporating area connectivity through time (TS) but dispersal is recovered by adding the parameter j (founder-event or long-distance dispersal speciation). In the second case, Lagrange infers

		Crown Calisto	Stem Cuban Calisto	Stem Jamaican Calisto	Stem Bahamian C. sibylla	Stem Bahamian C. apollinis	N° vicariant events	N° dispersal events
	NS0	PR-sH: 0.24 (-74.94);	nH-eC: 0.44 (-74.34);	*sH-Ja: 0.61 (-74.01);	eC-Ba: 0.55 (-74.12);	*eC-wC-Ba: 0.83 (-73.71)		
		PR-nH-sH: 0.16 (-75.36);	nH: 0.43 (-74.36)	sH: 0.27 (-74.83)	eC: 0.42 (-74.4)		8	5
		PR-nH: 0.11 (-75.7)						
Lagrange	NS1	*PR-nH: 0.48 (-74.62);	nH: 0.52 (-74.55);	sH-Ja: 0.42 (-74.75);	eC-Ba: 0.5 (-74.59);	*eC-wC-Ba: 0.81 (-74.1)	7	12 (1)
C++		PR-nH-sH: 0.16 (-75.73)	nH-eC: 0.44 (-74.71)	sH: 0.32 (-75.02)	eC: 0.48 (-74.62)		/	12 (1)
	TS2	PR-sH: 0.25 (-94.52);	*nH-eC: 0.95 (-93.18)	*sH-Ja: 0.97 (93.16)	*eC-Ba: 0.96 (-92.86)	*eC-wC-Ba: 0.95 (-92.86)		
		PR-nH-sH-eC: 0.22 (-94.6);					11	9
		PR-sH-eC: 0.22 (-94.6)						
	NS1-j	*PR-nH-sH: 0.92	*nH: 0.68;	sH: 0.49;	eC: 0.49;	Ba: 0.4;		
			eC: 0.29	Ja: 0.45	Ba: 0.48	eC: 0.2;	1	7 (6)
						wC: 0.2		
	NS1	*PR-nH-sH: 0.73;	*nH-eC: 0.52;	*sH-Ja: 0.72	*eC-Ba: 0.84	*eC-wC-Ba: 0.66;		
		PR-sH: 0.22	eC: 0.16			eC-Ba: 0.16;	10	3
BioGeoBEARS DEC model						wC-Ba: 0.16		
	TS1-j	*PR-sH: 0.72;	nH: 0.57;	*sH: 0.99	*eC: 0.81;	eC: 0.38;		
		PR-nH-sH: 0.12	eC: 0.37		wC: 0.18	wC: 0.28;	1	9 (6)
						eC-wC: 0.27		
	TS1	*PR-sH: 0.84	*nH-eC: 0.66;	*sH: 0.99	*eC: 0.65;	eC-wC-Ba: 0.45;	c	0 (E)
			nH: 0.3		wC: 0.21	eC-Ba: 0.43	6	9 (5)

Table 4 Biogeographical reconstructions for the evolution of Calisto

We excluded from the comparison the TS1 from Lagrange because of the unrealistic scenarios that were recovered (see text). Critical nodes for testing the Caribbean paleogeographical (vicariance) model are shown with their correspondent reconstructed ancestral geographical range. Preferred node distributions are highlighted in bold text and preceded by an asterisk (*). The number of well-supported vicariance and dispersal events were only counted when the relative probability of the best inference is two times larger than the following reconstructed distribution, in both immediate ancestral and daughter nodes. Dispersal events include the number of anagenetic range-switching and cladogenetic founder-events (the latter in parenthesis).



ancestral states by local optimization whereas BioGeo-BEARS reports ancestral states under the most likely model. This difference is evidenced in NS1 (analysis replicated using both software programs) where vicariance is only reconstructed under the global most-likely inference by BioGeoBEARS.

We believe, given the paleogeographic scenario and our dating estimations which correlate with the former, that the most plausible explanation for the colonization of Cuba is vicariance. Furthermore, the whole of extant Cuban diversity is monophyletic and sister to a Hispaniola lineage, as predicted by the vicariant model. Dispersal into Cuba 10–25 Ma, on the other hand, was not "long-distance" because both land blocks were quite close apart, if not physically connected. Thus, if dispersal were actually the main process, we would expect several independent Cuban lineages of varied ages surviving to the present (see extinction rates in "Diversification of *Calisto*" section) (Figure 2).

Vicariance driving speciation within islands is significantly recovered for Hispaniolan fauna during two instances, at 10–13 Ma and 4–6 Ma. The first vicariant instance is independently evidenced in two lineages with simultaneous shifts in ancestral ranges, the lyceius and the confusa-hysius clades. The dating estimates are congruent with the major uplift of the Cordillera Central which might have provided new ecological opportunities and created isolated populations [39,40]. Presence of local adaptations are evidenced not only by the disjunctive distributions of several sister-species pairs found on the northern/southern Hispaniola paleoislands respectively (e.g. C. tasajera and C. schwartzi are allopatrically adapted to mesophilic and forested montane habitats in Cordillera Central (nH) and Sierra de Bahoruco (sH) [46,55,56]), but also by ecological niche restrictions. For instance, sister species-pairs within both major clades feed, as larvae, exclusively on distinct bunch grasses, and have morphologically adapted to specific altitudinal ranges. Species inhabiting lower altitude and warmer areas are smaller than their sister montane species [27,46], suggesting an adaptation for thermoregulatory efficiency [57].

The second instance of vicariant process within Hispaniola occurred during the Pliocene as evidenced in the *lyceius*

Table 5 Diversification dynamics of Calisto as reconstructed by the R package laser

Diversification	rates for Ca	listo									
	LH	AIC	r ₁	r ₂	r ₃	а	Х	k	t_shift ₁	t_shift ₂	Δ AIC (yule-3-rates)
pb	-32.301	66.602	0.07	-	-	-	-	-	-	-	13.32
bd	-32.301	68.602	0.07	-	-	0	-	-	-	-	15.32
ddx	-28.705	61.41	0.394	-	-	-	0.599	-	-	-	8.124
*ddl	-26.579	57.158	0.175	-	-	-	-	40.119	-	-	3.871
spvar	-28.84	63.681	0.229	-	-	0.004	-	-	-	-	10.39
exvar	-32.36	70.721	0.069	-	-	0.014	-	-	-	-	17.43
bothvar	-28.826	65.653	0.227	-	-	0.004	-	-	-	-	12.37
yule-2-rates	-26.056	58.112	0.094	0.014	-	-	-	-	4.079	-	4.826
*yule-3-rates	-21.643	53.286	0.21	0.083	0.007	-	-	-	20.903	4.058	0
Diversification	rates for Ca	<i>listo</i> exclud	ling Cuba	n clade							
-	111					-	v	Ŀ	t chift	t chift	

	LH	AIC	r ₁	r ₂	r ₃	а	х	k	t_shift ₁	t_shift ₂	Δ AIC (ddl)
pb	-33.893	69.787	0.056	-	-	-	-	-	-	-	11.19
bd	-33.893	71.787	0.056	-	-	0	-	-	-	-	13.19
xbb	-29.428	62.855	0.55	-	-	-	0.889	-	-	-	4.258
*ddl	-27.299	58.597	0.186	-	-	-	-	24.646	-	-	0
pvar	-29.332	64.664	0.253	-	-	0.004	-	-	-	-	6.066
exvar	-33.951	73.902	0.055	-	-	0.018	-	-	-	-	15.3
oothvar	-29.323	66.646	0.251	-	-	0.004	-	-	-	-	8.048
yule-2-rates	-28.178	62.355	0.073	0.016	-	-	-	-	5.11	-	3.758
yule-3-rates	-25.313	60.626	0.21	0.055	0.008	-	-	-	20.903	5.079	2.029

The best models for our data is either DDL or yule-3-rates, which are highlighted in bold text and with an asterisk (*). Both models, nonetheless, predict a decreasing in diversification rates through time. Excluding the Cuban clade resulted in DDL, yule-2 or yule-3-rates as the main processes for the diversification of Hispaniolan lineages. LH: the best recovered log-likelihood, *r*; net diversification rate at time *i* ($\lambda_i - \mu_i$), *a*: extinction fraction (μ_i / λ_i), *X*: parameter controlling the magnitude of rates (only in DDL), *t*_shift; diversification shift at time *i*. The diversification models are pure-birth (pb), birth-death (bd), density-dependent speciation rate model following exponential (ddx) or logistic variants (ddl), exponential decline of speciation with constant extinction (spvar), exponential increase of extinction with constant speciation (exvar), speciation and extinction changes through time (bothvar), and pure birth models with *n* shifts in speciation (yule-n-rate).

and *chrysaoros* clades. Although an uplift of the Cordillera Central might have played a role in separating populations, the most likely explanation for the northern/southern paleoislands distributions might be related to the inundation of the Cul-de-Sac/Enriquillo depression, which acted as an effective barrier. Ecological niche shifts might be another plausible explanation for the *lyceus* clade members having differentiated during the Pliocene. As larvae, they feed on the bunchgrass *Uniola virgata*, which provides a unique niche and would have required significant adaptations [56].

The crown node ancestral distribution of Cuban and Bahamian *Calisto* is recovered as "eastern Cuba (eC)". Its sister taxa are Hispaniolan lineages that occur in the northwestern Cordillera Central (Massif du Nord in Haiti) [22,46], which is the closest region to eastern Cuba. Dispersal to central and western Cuba from "eC" appears to be the likeliest biogeographic scenario [28], although vicariance as the main process is only detected in NS1 from BioGeoBEARS. Dispersal dates are in line with the closure of the Havana-Matanzas Channel at 8–5 Ma, as well as with the accretion of Bahamian shallows and keys in the Pliocene/Pleistocene [6]. The two Bahamian lineages have distinct ancestral areas, while *C. sibylla* has an older origin and its source area is "eC", *C. apollinis* dispersed more recently from "western Cuba (wC)" (Figure 3).

Calisto diversification on the Greater Antilles and the Bahamas

The species richness of *Calisto* across islands is largely unequal. Such a pattern has been previously reported as the consequence of island size and age, ecological limits and habitat diversity [8,38,58]. Munroe [59,60] pointed out that extant *Calisto* diversity is distributed unequally among islands more likely due to speciation rather than to differential immigration, and that extinction was extremely low, especially in Hispaniola. The calculation of diversification rates and ancestral states in this study suggested that the extant geographical distribution of *Calisto* reflects the rapid diversification within Hispaniola

											Analy	ses acc	ount 8 m	nissing	spe	cies			
		λ_0	μ ₀	Ko	λ_1	μ	K ₁	t_shift	log-LH	Akaike weight	$\overline{\lambda_0}$	μo	Ko	λ1	μ	K ₁	t_shift	log-LH	Akaike weight
	Calisto diversification																		
CR0	constant λ and μ (birth-death)	0.07	0	-	-	-	-	-	-124.437	0.00	0.08	0	-	-	-	-	-	-123.853	0.00
CR1	λ declining as div-dep. No μ (DDL)	0.175	-	40.119	-	-	-	-	-118.715	0.03	-	-	-	-	-	-	-	-	-
CR2	div-dep with μ (DDL + E) depend in λ	0.163	0	43.055	-	-	-	-	-118.926	0.01	0.163	0	54.745	-	-	-	-	-118.774	0.02
SR0	Yule-2-rate	0.122	-	36.641	0.3	-	K ₀	13.567	-116.243	0.12	0.122	-	45.321	0.298	-	K1	13.567	-116.487	0.16
*SR1	shift in K	0.306	0	13.132	λο	μο	36.62	13.567	-113.877	0.48	0.295	0	14.456	λο	μ_0	45.466	13.567	-114.384	0.47
SR2	shift in K and $\boldsymbol{\mu}$	0.306	0	13.13	λ_0	0	36.617	13.567	-113.878	0.18	0.3	0.006	14.342	λ_0	0	45.355	13.567	-114.369	0.18
SR3	shift in K and $\boldsymbol{\lambda}$	0.322	0	13.025	0.298	μ_0	36.713	13.567	-113.863	0.18	0.289	0	14.507	0.298	μ_0	45.414	13.567	-114.382	0.17
KI1	shift in K in subclade	0.133	0	27.172	λ_0	μ	Inf.	13.567	-118.634	0.00	0.139	0	37.103	λ_0	μ	Inf.	13.567	-121.355	0.00
KI2	shift in K and $\boldsymbol{\mu}$ in subclade	0.133	0	27.159	λ_0	0	Inf.	13.567	-118.651	0.00	0.139	0	36.965	λ_0	0	Inf.	13.567	-121.371	0.00
KI3	shift in K and $\boldsymbol{\lambda}$ in subclade	0.164	0	25.591	0.115	μ_0	Inf.	13.567	-118.319	0.00	0.17	0	34.419	0.114	μ_0	Inf.	13.567	-120.853	0.00
KI4	shift in K, λ and μ in subclade	0.162	0	25.779	0.111	0	Inf.	13.567	-118.302	0.00	0.17	0	34.434	0.114	0	Inf.	13.567	-120.852	0.00
	Hispaniolan lineages diversification																		
CR0	constant λ and μ (birth-death)	0.056	0	-	-	-	-	-	-85.5	0.00	0.066	0	-	-	-	-	-	-84.936	0.00
CR1	λ declining as div-dep. No μ (DDL)	0.186	-	24.646	-	-	-	-	-78.905	0.19	-	-	-	-	-	-	-	-	-
CR2	div-dep with μ (DDL + E) depend in λ	0.165	0	26.665	-	-	-	-	-80.231	0.02	0.166	0	36.387	-	-	-	-	-79.968	0.04
SR0	Yule-2-rate	0.141	-	24	0.335	-	K ₀	12.843	-77.021	0.17	0.141	-	32	0.347	-	K1	12.843	-77.219	0.20
*SR1	shift in K	0.33	0.001	11.991	λο	μ_0	23.918	14.352	-75.372	0.32	0.339	0.001	15.518	λο	μ_0	31.942	12.843	-75.477	0.42
SR2	shift in K and $\boldsymbol{\mu}$	0.318	0.001	11.964	λ_0	0	24	14.352	-75.219	0.14	0.327	0	15.593	λ_0	0	32.023	12.844	-75.427	0.16
SR3	shift in K and $\boldsymbol{\lambda}$	0.333	0	12.98	0.245	μ	24.135	12.843	-75.09	0.16	0.295	0.001	15.743	0.355	μ	31.948	12.843	-75.372	0.17
	Cuban lineage diversification																		
CR0	constant λ and μ (birth-death)	0.09	0	-	-	-	-	-	-40.903	0.01	-	-	-	-	-	-	-	-	-
CR1	λ declining as div-dep. No μ (DDL)	0.27	-	14.568	-	-	-	-	-38.31	0.14	-	-	-	-	-	-	-	-	-
CR2	div-dep with μ (DDL + E) depend in λ	0.219	0	17.498	-	-	-	-	-39.235	0.02	-	-	-	-	-	-	-	-	-

Table 6 Diversification dynamics of *Calisto* as reconstructed by the R package *DDD*

Table 6 Diversification dynamics of Calisto as reconstructed by the R package DDD (Continued)

*SR0	Yule-2-rate	0.099	-	14	0.493	-	Ko	10.475	-35.409	0.35	-	-	-	-	 -	-	-	-
SR1	shift in K	0.502	0.012	2.323	λ_0	μ_0	13.644	11.894	-35.377	0.13	-	-	-	-	 -	-	-	-
*SR2	shift in K and $\boldsymbol{\mu}$	0.426	0.024	1.864	λο	0	13.982	13.567	-33.981	0.20	-	-	-	-	 -	-	-	-
SR3	shift in K and $\boldsymbol{\lambda}$	0.162	0.009	1.884	0.466	μο	13.718	13.573	-34.287	0.15	-	-	-	-	 -	-	-	-

The best models for each type of analyses, which include extinction and diversity-dependent processes, are highlighted in bold text and with an asterisk (*). λ is speciation rate, μ is extinction rate, K is species "carrying capacity" or a parameter analogous to it only in DDL. The estimated parameters to the right were calculated accounting missing taxa (8 species). The DDL model is not able to incorporate missing taxa whereas the Cuban clade in this study included all described species. A shift in K is recovered as the best explanation for the diversification patterns of *Calisto* and the Hispaniolan lineages. Yule-2-rate or a shift in K and μ are the best models to explain the diversification of the Cuban clade alone.

and Cuba during two instances, at 25 and 14 Ma, while inter-island flow was negligible for the entire genus.

Calisto is the most species-rich butterfly genus in the West Indies because it was able to expand its ecological niche (e.g. feeding on distinct bunchgrasses, tolerance to montane temperate and tropical conditions), which raised up the "ecological limits" on Calisto diversification. One sole change in the *K* parameter ("carrying-capacity for species diversity") is enough to explain the evolution of the whole genus. The recovered date of this shift at 14 Ma is congruent with an increase in ecological opportunity in Hispaniola and Cuba and a time at which new environments were being created as a result of geological processes (e.g. uplift of Cordilleras, unification of Cuban land blocks) [39,40]. The decoupling of clade "carrying capacity" and/or diversification rates of the Cuban lineage as the only explanation for the genus species richness is not supported. Nonetheless, the arrival of Calisto to an unoccupied island of Cuba did certainly provide for new heretofore empty niches to be colonized. The most likely scenario for such a decoupling was at 14 Ma, as recovered in DR1 analysis. However, because such a date is confounded with the availability of new niches in Hispaniola, a model including one single shift in K for the whole genus was preferred.

Adaptive radiation and the origin of island endemism of West Indies insects remain statistically untested. Under a phylogenetic framework, indirect evidence of adaptive radiation could be inferred based on diversification rate shifts: i.e. a rapid increase followed by a gradual reduction of diversification rate under a diversity-dependent process [61]. Calisto butterflies might have undergone two increases in diversification before they rapidly reached a "carrying-capacity" limit. The first one occurred during the uplift of Cordillera Central at 25 Ma (SR1 analysis), triggering a growth in Calisto diversification rate until all available niches were gradually occupied, at which time, probably, the speciation rate declined linearly with diversity (K = 12). It is unlikely that the extinction rate rose, as it was near zero in all of our estimations. The second major radiation took place at 14 Ma (discussed above), but it is more plausible that the diversification rate increased due to a shift in K rather than by a sole increase in speciation rate. Furthermore, the "Inf." values of K recovered for the Cuban clade in DR analyses might be an indication that the diversification rate has not yet reached its "ecological limit". The Cuban clade, when analyzed independently, better fits a 2-yule-rate, with 5 times larger speciation rate at 10 Ma than when the lineage branched off at 21 Ma.

An intriguing question is why the observed diversification dynamics of *Calisto* on Cuba and Hispaniola were not replicated on Jamaica and Puerto Rico, the third and fourth largest islands of the West Indies, respectively.

Whereas Calisto are usually locally adapted to particular habitats within Cuba and Hispaniola, the single species on each of the other two islands are widespread. While some diverse Hispaniolan lineages feed as larvae on bunch grasses, the Puerto Rican C. nubila is adapted to widespread-wide-blade grass feeding [56]. According to Turner, similar, relatively adaptable oviposition behaviour is exhibited by C. zangis of Jamaica [62]. Perhaps in this indiscriminate behaviour lies the explanation for the fact that these two species were able to colonize their entire respective islands instead of forming separate disjunctive populations as did their Hispaniolan congeners. Such wide distribution and relatively good dispersal abilities of these relatively larger Calisto species (Sourakov, pers. obs.) may have increased gene flow and hence prevented divergence. Further research on the natural history, dietary preferences and behaviour of Calisto is necessary to corroborate our speculations.

Conclusions

The phylogenetic and biogeographical evidence presented in this study agrees with the Caribbean paleogeographical model of colonization (Figures 2 and 3). Vicariant models explaining the diversification of Calisto have already been proposed based on their extant geographical distribution [31,33,36,60], although some authors had favoured the alternative dispersalist explanation [27,63]. Here we observed that the evolution of Calisto passed through both vicariant processes and long-distance dispersals. However, the most important means for diversity origination in this largest genus of West Indies butterflies, was intra-island rapid radiation through key innovations (e.g. unusual larval hostplant, adaptation to montane, temperate and tropical conditions) and the availability of ecological niches triggered by environmental changes (e.g. accretion of mountain ranges, different island configuration and areaconnectivity through time). Nonetheless, more rigorous tests and associations between ecological niche spectrum, phenotypic variability and selection within these butterflies are needed to give the adequate weight to abiotic factors (geographic and climatic) and niche specializations in the observed burst followed by a slowdown in diversification rates.

Methods

Taxon sampling

We included 36 out of the 44 described *Calisto* species (Additional file 2). Species sampling took place across the entire geographical distribution of the genus in the Greater Antilles, except for the Anegada Island where only one species occurs. Our analyses also included DNA sequences previously reported from taxa across the tribe Satyrini and *Calisto* [27-29,32] (Additional file 2). Species identifications were based on morphology and the DNA

barcode region was used for further corroboration [64]. Voucher photographs are available at the Nymphalidae Systematics Group (NSG) Voucher Database (nymphalidae. utu.fi) and in BOLD (boldsystems.org).

Dataset acquisition

Genomic DNA was isolated from two butterfly legs using the QIAGEN's DNeasy kit. We used sequences of six standard molecular markers for nymphalid butterflies, one mitochondrial – COI (1487 bp) – and five nuclear genes – CAD (850 bp), EF-1 α (1240 bp), GAPDH (691 bp), RpS5 (617) and *wingless* (400 bp). Primer pair sequences and laboratory protocols are described in [65]. DNA Sanger sequencing was carried out by the company Macrogen and each gene sequence was edited and manually aligned using the program BioEdit v7.0.5 [66]. Datasets were generated in different input formats using the web application VoSeq v1.7.0 [67].

Phylogenetic analyses

We used single-gene and combined datasets. We partitioned our single-gene datasets by codon position and our combined dataset by gene sequences in all analyses. In addition we used character groupings of similar relative evolutionary rates as an alternative strategy for Bayesian Inference (BI) [68], after determining that the gene trees were not in conflict with each other. We used the software TIGER [69] to subdivide our combined dataset into 12 "bins" each containing a number of characters with similar relative rates: bin1 = 2739, bin2 to bin5 = 0, bin6 = 12, bin7 = 7, bin8 = 21, bin9 = 87, bin10 = 525, bin11 = 1269 and bin12 = 637. We combined the "bins" that contained fewer than 500 sites (bin2 to bin9) with the invariable bin1, resulting in four character groupings which were used for our alternative partitioning approach (Table 1).

We performed 1000 Maximum Likelihood (ML) pseudo-replicates analyses using RaXML v7.3.1 [70] on the Bioportal server [71], selecting the thorough bootstrap algorithm and the mix option for the evolutionary model. The BI analyses were carried out using MrBayes v3.2.1 [72] on the Bioportal server. We performed 10 million generations with sampling every 1000 generation and four chains, one cold and three heated, for two independent runs. The parameters and models of evolution were unlinked across character partitions. We selected the mixed evolutionary model option in all BI analyses whereas in the alternative partitioning strategy, we selected the corresponding model for each "bin" as calculated in jModelTest 0.1.1 [41] based on Bayesian Information Criterion (BIC). The convergence of the two runs on each BI was ascertained by visual inspection of the log-likelihoods stationary distribution, discarding the first 25% sampled trees, as well as by checking that the final average standard deviation of split frequencies was below 0.05 and that the potential scale reduction factor (PSRF) for each parameter was close to 1.

Time of diversification estimates

Because there is no fossil record reported for the genus, we reconstructed a broader phylogeny including most of the representatives of the Satyrini subtribes that are closely related to Calisto [32] (Additional file 2) and constrained it with secondary calibration points from a fossil-calibrated Nymphalidae phylogeny [73]. We selected only one terminal per Calisto species to maximize the gene coverage in the resulting dataset. We also made an analysis excluding the genus Euptychia because long branch attraction affecting the position of Calisto has been reported [32]. The selected calibration points were chosen from well-supported monophyletic groups: the root of the tree to 49.1 ± 5 Ma, the crown age of the tribe Satyrina to 24.7 ± 4 Ma and the crown age of Euptychiina excluding Euptychia, Paramacera and Cyllopsis to 35.1 ± 4 Ma.

The dating analyses were run in BEAST v1.7.4 [74] and executed on the Bioportal server. We partitioned our dataset by gene sequence and set the corresponding substitution model as calculated in jModelTest (Table 1) and the uncorrelated log-normal relaxed clock model for each partition. We applied either the Birth-Death or the Calibrated Yule speciation processes as the tree prior in separate analyses to investigate the impact of this parameter on the final age estimates. In addition, the calibration points were modelled as either normal distributions (soft bounds) or uniform ranges (hard bounds). Finally, we set the mean rate of the molecular clock (ucld prior) with a uniform distribution between 0.0 and 10.0 and left other priors as default.

Each analysis was run four independent times for 50 million generations each and sampling trees and parameters every 5000th generation. We discarded the first 2500 sampled trees from each run as burnin. We verified in Tracer v1.5 the convergence and good mixing of MCMC as well as the Effective Sample Size of each estimated parameter to be higher than 200. Output .log and .tre files were combined in LogCombiner v1.7.4 after resampling a third of the post-burnin trees from each run. Trees were summarized in TreeAnnotator v1.7.4 into a single maximum clade credibility tree with node information calculated as mean heights.

Historical biogeography reconstruction

We used our dated chronogram for *Calisto* as the input tree, excluding outgroups and *C. pulchella* because its distribution has been altered by sugar cane introduction, on which it is currently a pest [33]. The following subdivision of areas was set: "PR" – Puerto Rico; "nH" – the northern Hispaniola paleoisland, including Cordillera Central/Massif du Nord, Sierra de Neiba/Chaîne des Matheux, and eastern Hispaniola; "sH" – the southern Hispaniola paleoisland, including Sierra de Bahoruco/ Chaîne de la Selle and Massif de la Hotte in Tiburón Peninsula; "Ja" – Jamaica; "eC" – the eastern Cuba, including Nipe – Sagua – Baraoca and Sierra Maestra mountain ranges; "wC" – the central and western Cuba, including Guamuhaya and Guaniguanico mountain ranges; "Ba" – the Bahamas. Distributional ranges of *Calisto* were taken from several sources [22,27-29,33,46].

We used the Dispersal-Extinction-Cladogenesis (DEC) model as implemented in Lagrange C_{++} [75,76]. DEC is a realistic and flexible model for biogeographical reconstructions that estimates the probabilities (likelihoods) of ancestral geographical distributions, and it allows the parameterization of dispersal through time according to the geological history of a region. We conducted analyses using a non-time-stratified approach (NS) and different dispersal rates across time slices (stratified, TS). The nontime-stratified analysis NS0 was conducted under default settings. The maximum distributional range was constrained to three areas and we excluded distributions with unlikely area-connectivity (e.g. Puerto Rico and Western Cuba) in NS1 analysis. TS1 used four time slices, subdividing the phylogeny at 5 Ma, 10 Ma and 20 Ma. Dispersal rate matrices were constructed according to the paleogeographical configuration on each time slice. Probabilities to disperse were set to 0.75 when two areas were adjacent, to 0.5 when two areas were weakly separated by a geographical barrier (e.g. the Cul-de-Sac/ Enriquillo depression), to 0.1 when two areas were separated by water of a distance less than 200 km (e.g. northern Hispaniola and eastern Cuba), to 0.01 for longdistance dispersal, including one extra area and/or >200 km water-crossing (e.g. Puerto Rico to southern Hispaniola), and to 0.0001 for other kinds of long-distance dispersal.

We found a particular node in TS1 analysis to be unlikely (the Cuban-Bahamian subclade including *C. sibylla* and *C. apollinis*). This group had a crown age of $10 \pm$ 2 Ma and an ancestral range eC-Ba after TS1. Paleogeographically, this is improbable because the Bahamas were submerged at least until the Pliocene (~5 Ma). We thus constrained such node to "eC" in TS2 because Lagrange, as it is currently implemented, does not allow the exclusion of unlikely area-connectivity through time slices.

Moreover, several sets of area distribution were independently constrained at the root of the *Calisto* tree to maximize the global likelihood of NS1 and TS2 and to compare the statistical support of likely ancestral ranges. We also used the R package BioGeoBEARS [77,78] which implements the DEC model similar to Lagrange C++ but with the possibility of increasing the number of free parameters. We allowed the founder-event speciation parameter *j*

to be estimated in NS1-*j* and TS1-*j* to evaluate the importance of long-distance dispersal across islands. Another advantage of BioGeoBEARS is that distinct area-connection through time is allowed, hence we created an areaconnectivity matrix for each time slice in TS1 and TS1-*j* (Figure 3).

Diversification of Calisto

We used the packages *laser* [79], *ape* [80] and *DDD* [61,81] in R [82] to investigate the mode of diversification of extant *Calisto* taxa. Lineage Through Time (LTT) plots with confidence intervals representing a pure-birth null hypothesis model were made using *ape*. We compared different models of cladogenesis allowing temporal shifts in diversification rates using the Akaike Information Criterion differentials ($\Delta AIC_{RC} = AIC_{RC}$ (best rate-constant model) - AIC_{RV} (best rate-variable model)) as implemented in *laser*. We also computed the ΔAIC_{RC} separately for the *Calisto* phylogeny, excluding the Cuban species.

We used the R package *DDD* to fit the best phylogenetic diversification model that would explain the evolutionary history of Calisto. The analyses included three main models: a constant-rate evolution (CR), a shift in net diversification rate at some point in time (SR) and a decoupling of rates between the Cuban clade and the remaining taxa (DR). CR models incorporated either constant birth-death process (CR0), a decrease in speciation rate following a density-dependent process without extinction (CR1) or a decrease in speciation rate following a diversity-dependent process, including the estimation of extinction rate (CR2). SR models were set up to: one shift in speciation rate (yule-2-rate model) (SR0), one shift in species carrying capacity K (SR1), one shift in K and extinction rate (SR2), or one shift in K and speciation rate (SR3). DR models described one single shift in *K* for the Cuban clade (DR1), one shift in K and extinction rate for the Cuban clade (DR2), one shift in K and speciation rate for the Cuban group (DR3), and one shift in K, speciation and extinction rates for Cuban taxa (DR4). Moreover, we conducted CR and SR analyses for the main Calisto tree, excluding the Cuban clade, and for the Cuban clade independently. Comparisons between different phylogenetic diversification models were done using Akaike weights.

Availability of supporting data

The data sets supporting the results of this article are available in the TreeBASE repository, in http://purl.org/phylo/treebase/phylows/study/TB2:S16186?format=html [83].

Additional files

Additional file 1: Additional phylogenetic trees of *Calisto*. In order of appearance in the pdf file: gene trees using COI (page 1), CAD (page 2), EF1a (page 3), GAPDH (page 4), RpS5 (page 5) and wingless (page 6) single

gene datasets in MrBayes. Each node displays posterior probabilities values. Calibrated *Calisto* trees as estimated using BEAST including the genus *Euptychia* and using priors on calibration points and speciation process respectively: Normal and Birth-Death (BD) (page 7), Normal and Yule (page 8), Uniform and BD (page 9), Uniform and Yule (page 10). Calibrated *Calisto* trees excluding the genus *Euptychia* and using: Normal and BD (page 11), Normal and Yule (page 12), Uniform and BD (page 13), Uniform and Yule (page 14). On the calibrated trees, posterior probabilities are displayed on each node, 95% confidence interval for the dating estimates are shown as bars on each node, two red stars on every tree show the calibration points that were used in the present study and the y-axis represents time in million years. Subtribes names are represented on the left of each clade.

Additional file 2: List of specimens and voucher information used in the present study. A. xls file displaying each sequenced gene are presented with their corresponding GenBank accession number, unless the sequence is not available in the database where a "X" is shown. The hyphen "-" indicates unsuccessful DNA sequencing for that particular gene.

Abbreviations

Ma: Million years ago; AlC: Akaike information criterion; DEC: Dispersal-Extinction-Cladogenesis model; Bl: Bayesian inference; ML: Maximum likelihood.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

NW and PMM conceived the initial research plan, AS, CP, JYM, RNA and NW collected and provided the specimens for this study; PMM, RNA and CP did the majority of the laboratory work and PMM, most of the data analyses. PMM wrote the first draft of the manuscript and the rest of the authors contributed actively to the final version. All authors read and approved the final manuscript.

Acknowledgements

We thank A.V.Z. Brower, F.L. Condamine, Alexandra Sourakov and an anonymous reviewer for their constructive comments on an early version of this study. We are also grateful to NJ. Matzke, R.S. Etienne and J. Smrčková for their advice on the biogeographical and diversification rate analyses. We thank Kelvin Guerrero for help with acquiring permits and collecting samples in the Dominican Republic. The National Geographic Society funded some of the fieldwork through Committee for Research and Exploration grant #5717-96. The Kone Foundation is thanked for funds used in laboratory work. P.M.M. acknowledges the funds from Oskar Öflunds Foundation, Turun yliopistosäätiö and GAJU grant 156/2013/P. Thomas Turner is acknowledged for contributing a key specimen used in the DNA analysis.

Author details

¹Laboratory of Genetics, Department of Biology, University of Turku, FI-20014 Turku, Finland. ²School of Biological Sciences, University of South Bohemia and Institute of Entomology, Biology Centre AS CR, CZ-37005 Ceske Budejovice, Czech Republic. ³División de Colecciones Zoológicas y Sistemática, Instituto de Ecología y Sistemática, Carretera de Varona km 3.5, Capdevila, Boyeros, Ciudad de La Habana, Cuba. ⁴McGuire Center for Lepidoptera and Biodiversity, Florida Museum of Natural History, University of Florida, Gainesville, FL 32611, USA.

Received: 8 May 2014 Accepted: 2 September 2014 Published online: 16 September 2014

References

- 1. Darlington PJ: The origin of the fauna of the Greater Antilles, with discussion of dispersal of animals over water and through the air. *Q Rev Biol* 1938, 13:274–300.
- Hedges SB, Hass CA, Maxson LR: Caribbean biogeography: molecular evidence for dispersal in West Indian terrestrial vertebrates. Proc Natl Acad Sci U S A 1992, 89:1909–1913.
- Hedges SB: Historical biogeography of West Indian vertebrates. Annu Rev Ecol Syst 1996, 27:163–196.
- Dávalos LM: Phylogeny and biogeography of Caribbean mammals. Biol J Linn Soc 2004, 81:373–394.

- Rosen DE: A vicariance model of Caribbean biogeography. Syst Zool 1975, 24:431–464.
- Iturralde-Vinent MA: Meso-Cenozoic Caribbean paleogeography: implications for the historical biogeography of the region. Int Geol Rev 2006, 48:791–827.
- Ricklefs RE, Cox GW: Taxon cycles in the West Indian avifauna. Am Nat 1972, 106:195–219.
- Losos JB, Schluter D: Analysis of an evolutionary species-area relationship. Nature 2000, 408:847–850.
- Alonso R, Crawford AJ, Bermingham E: Molecular phylogeny of an endemic radiation of Cuban toads (Bufonidae: *Peltophryne*) based on mitochondrial and nuclear genes. J Biogeogr 2012, 39:434–451.
- Ali JR: Colonizing the Caribbean: is the GAARlandia land-bridge hypothesis gaining a foothold? *J Biogeogr* 2012, 39:431–433.
- Van Gestel J-P, Mann P, Grindlay NR, Dolan JF: Three-phase tectonic evolution of the northern margin of Puerto Rico as inferred from an integration of seismic reflection, well, and outcrop data. *Mar Geol* 1999, 161:257–286.
- MacPhee RDE, Iturralde-Vinent MA, Gaffney ES: Domo de Zaza, an early Miocene vertebrate locality in south-central Cuba, with notes on the tectonic evolution of Puerto Rico and the Mona Passage. Am Museum Novit 2003, 3394:1–42.
- Pindell JL, Barrett SF: Geological evolution of the Caribbean Region: a plate-tectonic perspective. In Geol North Am Vol H, Caribb Reg. Edited by Dengo G, Case JE. Boulder, CO: Geological Society of America; 1990:405–432.
- Hedges SB: Biogeography of the West Indies: an overview. In *Biogeogr* West Indies Patterns Perspect. Edited by Woods CA, Sergile FE. Boca Raton: CRC Press; 2001:15–33.
- 15. Graham A: Models and Cenozoic paleoenvironments of the Caribbean region. Syst Bot 2003, 28:378–386.
- Lewis JF, Draper G: Geology and tectonic evolution of the Northern Caribbean Margin. In Geol North Am Vol H, Caribb Reg. Edited by Dengo G, Case JE. Boulder, CO: Geological Society of America; 1990:77–140.
- Heubeck C, Mann P: Structural geology and cenozoic tectonic history of the Southeastern termination of the Cordillera Central, Dominican Republic. In *Geol Tecton Dev North Am - Caribb plate Bound Hisp.* Edited by Mann P, Draper G, Lewis JF. Geological Society of America Special Paper 262; 1991:315–336.
- Maurrasse FJ-MR, Pierre-Louis F, Rigaud J-G: Cenozoic facies distribution in the Southern Peninsula of Haiti and the Barahona Peninsula, Dominican Republic. Trans 9th Caribb Geol Conf 1980, 1:161–174.
- Robinson E: Jamaica. In *Caribb Geol An Introd*. Edited by Donovan SK, Jackson TA. Kingston, Jamaica: University of the West Indies Publishers Association; 1994:111–127.
- Pindell J: Evolution of the Gulf of Mexico and the Caribbean. In Caribb Geol An Introd. Edited by Donovan SK, Jackson TA. Kingston, Jamaica: The University of West Indies Publishers Association; 1994:13–40.
- Hearty PJ, Neumann AC: Rapid sea level and climate change at the close of the Last Interglaciation (MIS 5e): evidence from the Bahama Islands. *Quat Sci Rev* 2001, 20:1881–1895.
- 22. Michener CD: A review of the genus *Calisto* (Lepidoptera, Satyrinae). *Am Museum Novit* 1943, **1236**:1–6.
- Lamas G: Atlas of Neotropical Lepidoptera: Checklist: Part 4A. Hesperioidea-Papilionoidea, Volume 5A. Gainesville, Florida: Association for Tropical Lepidoptera; 2004. xxxvi + 439.
- 24. Scott JA: Biogeography of Antillean butterflies. Biotropica 1972, 4:32-45.
- Fontenla JL: Biogeography of Antillean butterflies (Lepidoptera: Rhopalocera): patterns of association among areas of endemism. *Trans Am Entomol Soc* 2003, **129**:399–410.
- Smith DS, Miller LD, KcKenzie F: The butterflies of Anegada, British Virgin Islands, with descriptions of a new *Calisto* (Satyridae) and a new *Copaeodes* (Hesperiidae) endemic to the island. *Bull Allyn Mus* 1991, 133:1–25.
- Sourakov A, Zakharov EV: "Darwin's butterflies"? DNA barcoding and the radiation of the endemic Caribbean butterfly genus *Calisto* (Lepidoptera, Nymphalidae, Satyrinae). *Comp Cytogenet* 2011, 5:191–210.
- Núñez Aguila R, Oliva Plasencia E, Matos Maravi PF, Wahlberg N: Cuban Calisto (Lepidoptera, Nymphalidae, Satyrinae), a review based on morphological and DNA data. Zookeys 2012, 165:57–105.
- Núñez Aguila R, Matos-Maraví PF, Wahlberg N: New Calisto species from Cuba, with insights on the relationships of Cuban and Bahamian taxa (Lepidoptera, Nymphalidae, Satyrinae). Zootaxa 2013, 3669:503–521.

- Miller LD: The higher classification, phylogeny and zoogeography of the Satyridae (Lepidoptera). Mem Am Entomol Soc 1968, 24:1–174.
- 31. Viloria AL: Historical biogeography and the origins of the satyrine butterflies of the tropical andes (Lepidoptera: Rhopalocera). In *Una Perspect Latinoam la Biogeogr.* Edited by Morrone JJ, Llorente J. México: Universidad Nacional Autónoma de México; 2003:247–261.
- Peña C, Nylin S, Wahlberg N: The radiation of Satyrini butterflies (Nymphalidae: Satyrinae): a challenge for phylogenetic methods. Zool J Linn Soc 2011, 161:64–87.
- Smith DS, Miller LD, Miller JY: The Butterflies of the West Indies and South Florida. Oxford: Oxford University Press; 1994:264.
- 34. Riley ND: A Field Guide to the Butterflies of the West Indies. London: Collins; 1975:224.
- Brown FM: The origins of the West Indian butterfly fauna. In Zoogeography Caribb. Edited by Gill FB. Pennsylvania: Academy of Natural Sciences of Philadelphia, Special Publication No. 13; 1978:5–30.
- Miller JY, Miller LD: The biogeography of the West Indian butterflies (Lepidoptera): an application of a vicariance/dispersalist model. In *Biogeogr West Indies Patterns Perspect.* 2nd edition. Edited by Woods CA, Sergile FE. Florida: CRC Press LLC; 2001:127–155.
- Miller LD, Miller JY: The Biogeography of the West Indian Butterflies (Lepidoptera: Papilionidea, Hesperioidea): a Vicariance model. In *Biogeogr West Indies Past, Present Futur.* Edited by Woods CA. Gainesville, Florida: Sandhill Crane Press; 1989:229–262.
- Rabosky DL, Glor RE: Equilibrium speciation dynamics in a model adaptive radiation of island lizards. Proc Natl Acad Sci U S A 2010, 107:22178–22183.
- Sly ND, Townsend AK, Rimmer CC, Townsend JM, Latta SC, Lovette IJ: Ancient islands and modern invasions: disparate phylogeographic histories among Hispaniola's endemic birds. *Mol Ecol* 2011, 20:5012–5024.
- Oneal E, Otte D, Knowles LL: Testing for biogeographic mechanisms promoting divergence in Caribbean crickets (genus Amphiacusta). *J Biogeogr* 2010, 37:530–540.
- Posada D: jModelTest: phylogenetic model averaging. Mol Biol Evol 2008, 25:1253–1256.
- Johnson K, Quinter EL, Matusik D: A new species of *Calisto* from Hispaniola with a review of the female genitalia of Hispaniolan congeners (Satyridae). J Res Lepid 1987, 25:73–82.
- Schwartz A: A new species of Calisto from the Cordillera Central, Republica Dominicana. Milwaukee Public Museum, Contrib to Biol Geol 1991, 81:1–3.
- Schwartz A, Gonzalez FL: A new species of Calisto (Satyridae) from Hispaniola. Bull Allyn Mus 1988, 117:1–5.
- Johnson K, Hedges SB: Three new species of Calisto from Southwestern Haiti (Lepidoptera: Nymphalidae: Satyrinae). Trop Lepid 1998, 9:45–53.
- Schwartz A: The Butterflies of Hispaniola. Gainesville: University Presses of Florida; 1989:583.
- 47. Rabosky DL: Likelihood methods for detecting temporal shifts in diversification rates. *Evolution (N Y)* 2006, **60**:1152–1164.
- Peña C, Nylin S, Freitas AVL, Wahlberg N: Biogeographic history of the butterfly subtribe Euptychiina (Lepidoptera, Nymphalidae, Satyrinae). Zool Scr 2010, 39:243–258.
- Matos-Maraví PF, Peña C, Willmott KR, Freitas AVL, Wahlberg N: Systematics and evolutionary history of butterflies in the "Taygetis clade" (Nymphalidae: Satyrinae: Euptychiina): towards a better understanding of Neotropical biogeography. Mol Phylogenet Evol 2013, 66:54–68.
- Elias M, Joron M, Willmott K, Silva-Brandão KL, Kaiser V, Arias CF, Gomez Piñerez LM, Uribe S, Brower AVZ, Freitas AVL, Jiggins CD: Out of the Andes: patterns of diversification in clearwing butterflies. *Mol Ecol* 2009, 18:1716–1729.
- Kodandaramaiah U, Wahlberg N: Phylogeny and biogeography of Coenonympha butterflies (Nymphalidae: Satyrinae) - patterns of colonization in the Holarctic. Syst Entomol 2009, 34:315–323.
- Wahlberg N, Freitas AVL: Colonization of and radiation in South America by butterflies in the subtribe Phyciodina (Lepidoptera: Nymphalidae). Mol Phylogenet Evol 2007, 44:1257–1272.
- Condamine FL, Silva-Brandão KL, Kergoat GJ, Sperling FAH: Biogeographic and diversification patterns of Neotropical Troidini butterflies (Papilionidae) support a museum model of diversity dynamics for Amazonia. BMC Evol Biol 2012, 12:82.
- 54. Fox RM: Affinities and distribution of Antillean Ithomiidae. *J Res Lepid* 1963, **2:**173–184.

- Hedges SB, Johnson K: Calisto tasajera in the Hispaniolan Cordillera Central (Lepidoptera: Nymphalidae: Satyrinae). Trop Lepid 1994, 5:93–94.
- Sourakov A: Notes on the genus *Calisto*, with descriptions of the immature stages (Part II) (Lepidoptera: Nymphalidae: Satyrinae). *Trop Lepid* 2000, 10:73–79.
- Shelly TE, Ludwig D: Thermoregulatory behavior of the butterfly *Calisto* nubila (Satyridae) in a Puerto Rican forest. Oikos 1985, 44:229–233.
- MacArthur RH, Wilson EO: The Theory of Island Biogeography. Princeton, NJ: Princeton University Press; 1967:203.
- Munroe EG: The Geographical Distribution of Butterflies in the West Indies. Ithaca, New York, USA: Cornell University; 1948:1–1100.
- 60. Munroe EG: The systematics of *Calisto* (Lepidoptera, Satyrinae), with remarks on the evolutionary and zoogeographic significance of the genus. *J New York Entomol Soc* 1950, **58**:211–240.
- Etienne RS, Haegeman B: A conceptual and statistical framework for adaptive radiations with a key role for diversity dependence. *Am Nat* 2012, 180:E75–E89.
- 62. Brown FM, Heineman B: Jamaica and its Butterflies. London: E. W. Classey; 1972:478+xv.
- Wetherbee DK: Eighth Contribution on the Larvae And/or Larval Host Plants of Hispaniolan Butterflies: The Butterfly/larva Flora with a Theory of Calisto (Satyridae) Origin and Endemicity. Shelburne, Massachsetts: Selfpublished; 1992:1–173.
- 64. Hebert PDN, Cywinska A, Ball SL, DeWaard JR: Biological identifications through DNA barcodes. Proc R Soc B Biol Sci 2003, 270:313–321.
- 65. Wahlberg N, Wheat CW: Genomic outposts serve the phylogenomic pioneers: designing novel nuclear markers for genomic DNA extractions of Lepidoptera. *Syst Biol* 2008, **57**:231–242.
- Hall TA: BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symp Ser 1999, 41:95–98.
- 67. Peña C, Malm T: VoSeq: a voucher and DNA sequence web application. PLoS ONE 2012, 7:e39071.
- Rota J, Wahlberg N: Exploration of data partitioning in an eight-gene dataset: phylogeny of metalmark moths (Lepidoptera, Choreutidae). Zool Scr 2012, 41:536–546.
- 69. Cummins CA, McInerney JO: A method for inferring the rate of evolution of homologous characters that can potentially improve phylogenetic inference, resolve deep divergence and correct systematic biases. *Syst Biol* 2011, **60**:833–844.
- Stamatakis A: RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 2006, 22:2688–2690.
- Kumar S, Skjæveland Å, Orr RJS, Enger P, Ruden T, Mevik B-H, Burki F, Botnen A, Shalchian-Tabrizi K: AIR: a batch-oriented web program package for construction of supermatrices ready for phylogenomic analyses. BMC Bioinformatics 2009, 10:357.
- Ronquist F, Teslenko M, Van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP: MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol 2012, 61:539–542.
- Wahlberg N, Leneveu J, Kodandaramaiah U, Peña C, Nylin S, Freitas AVL, Brower AVZ: Nymphalid butterflies diversify following near demise at the Cretaceous/Tertiary boundary. Proc R Soc B Biol Sci 2009, 276:4295–4302.
- 74. Drummond AJ, Suchard MA, Xie D, Rambaut A: Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Mol Biol Evol* 2012, 29:1969–1973.
- 75. Ree RH, Smith SA: Maximum likelihood inference of geographic range evolution by dispersal, local extinction, and cladogenesis. *Syst Biol* 2008, 57:4–14.
- 76. Smith SA: Taking into account phylogenetic and divergence-time uncertainty in a parametric biogeographical analysis of the Northern Hemisphere plant clade Caprifolieae. *J Biogeogr* 2009, **36**:2324–2337.
- Matzke NJ: BioGeoBEARS: BioGeography with Bayesian (and Likelihood) Evolutionary Analysis in R Scripts. 2013. http://cran.r-project.org/web/ packages/BioGeoBEARS/index.html.
- Matzke NJ: Probabilistic historical biogeography: new models for founder-event speciation, imperfect detection, and fossils allow improved accuracy and model-testing. *Front Biogeogr* 2013, 5:242–248.
- Rabosky DL: LASER: a maximum likelihood toolkit for detecting temporal shifts in diversification rates from molecular phylogenies. *Bioinformatics* 2006, 2:247–250.

- 80. Paradis E, Claude J, Strimmer K: APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* 2004, **20**:289–290.
- 81. Etienne RS, Haegeman B: DDD: Diversity-Dependent Diversification. 2013. http://cran.r-project.org/web/packages/DDD/index.html.
- 82. R Core Team: **R: A language and environment for statistical computing.** 2013. www.r-project.org.
- Matos-Maraví P, Núñez Águila R, Peña C, Miller JY, Sourakov A, Wahlberg N: DNA sequence data from the butterfly genus Calisto (Nymphalidae: Satyrinae: Satyrini). TreeBASE database. 2014. http://purl.org/phylo/treebase/phylows/ study/TB2:S16186?format=html.

doi:10.1186/s12862-014-0199-7

Cite this article as: Matos-Maraví *et al.*: **Causes of endemic radiation in** the Caribbean: evidence from the historical biogeography and diversification of the butterfly genus *Calisto* (Nymphalidae: Satyrinae: Satyrini). *BMC Evolutionary Biology* 2014 14:199.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

BioMed Central

(

Submit your manuscript at www.biomedcentral.com/submit